

Chemical variability of *Vicia* L. seed oils: incidence on phylogenetic relationships

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Abstract

Legume seeds are known to be a source of protein, vitamins, essential organic minerals and fatty acids. Their high protein content makes them a valuable natural and inexpensive alternative to the soybean. Seeds fatty acids of genotypes belonging to nine *Vicia* L. taxa collected in Algeria were extracted by Soxhlet and their composition was determined by Gas Chromatography coupled with Flame Ionization Detection (GC-FID). In order to assess phylogenetic relationships between taxa on intra and interspecific levels, fatty acids profiles were used as a chemotaxonomic marker. Oil yields were between 0.67 and 2.40%. A total of 27 fatty acids were identified, varying from 9 to 20 compounds. The proportion of unsaturated fatty acids varies from 70.67 to 91.749% where the linoleic acid is predominant. While the saturated fatty acids content is between 8.60 and 29.33% with the predominance of palmitic acid. Our results demonstrate the quality of the genotypes, which contain fatty acids of great nutritional interests. Indeed, Omega 3- fatty acids, which rate can reach 12.187% in our taxa, are Poly Unsaturated Fatty Acids associated with many health benefits, such as cardiovascular ones. Indeed, the results demonstrate the nutritional quality of obtained oils and opens up perspectives to explore other compounds. Finally, the hierarchical classification revealed two major clusters where the taxonomic boundaries of the genus are well defined at subgeneric and sectional levels. It is also an important step to suggest the species as a new industrial crop for the animal feed and production of proteins and oils widely used in food industries

Keywords: Algeria; Fatty acids; *Vicia*; GC-FID; Phylogeny.

Introduction

Legume seeds are recognized to be a rich source of protein, vitamins, essential organic minerals and fatty acids (FA) (Pastor Cavada et al. 2009). Their high protein content makes them a valuable natural and inexpensive alternative to the soybean. The widespread use of vetch seeds also makes them an important source of fat for ruminant and non-ruminant diet. The importance of FA lies in their role as a precursor for the biosynthesis of eicosanoids which is an important bio-regulator for many cellular metabolic processes. It is well established that CLA or other PUSFA increase pregnancy rates of cows. Milk and meat that contain high quantities of CLA can benefit for human health (Belury 2002).

In addition, anti-glycemic and anti-cholesterolemic legume seeds effects are due to dietary fiber and FA content (Pirman and Stibilj 2003). Therefore, FA attracted attention for their value for industrial purposes and also for their use as chemotaxonomic significance.

In Algeria, the genus *Vicia* is distributed in the whole country and is represented by 26 species. It includes many species and subspecies with morphological characteristics that are so subtly different which makes difficult to discriminate taxa within the genus. Despite ongoing researches, the few existing reports on *Vicia* L. taxa were conducted either on morphological or on molecular aspects, which did not cover the entire classification of the species. Therefore, chemotaxonomy based on Fatty Acids Methyl Esters could help to resolve uncertainties not completely addressed by other approaches, such as molecular studies. As far as we know, there are no previous studies on the intraspecific chemical variability of Algerian *Vicia* L. taxa fatty acids oils and their nutritional values, although its place as animal feed in the agricultural areas of the arid regions of the Mediterranean countries is well demonstrated in the literature.

The aim of our study is to assess the chemovariability of FA profiling of *Vicia* L. taxa. This work, combined with those we carried out previously on the genetic diversity of *Vicia* species publications (Bechkri and Khelifi 2016, Bechkri and Khelifi 2017 a, b, Bechkri *et al.* 2018, Bechkri *et al.* 2019), would furnish complementary insight to envisage the management, selection and propagation of interesting chemotypes and genotypes. It is also an important step to suggest the species as a new industrial crop for the animal feed and production of proteins and oils widely used in food industries.

In the same time, the FA profiles were used as chemotaxonomical markers to establish phylogenetic relationships between the taxa at different taxonomic levels, as we mentioned in our previous publications, the taxonomic boundaries within the genus are controversial.

Material and methods

Vicia seeds

Twenty vetch populations randomly collected from their natural habitats in various bioclimatic conditions of Algeria were used in the current study. Origins and field information are given in Table 1. Taxonomic identification of accessions was verified by the morphology of plants grown from seeds as it is explained in Bechkri and Khelifi (2016).

Table 1. Passport data of investigated taxa

Taxon	Code	Date of collection	Origin	Altitude (m)
<i>V. sativa subsp. consobrina</i>	14	1.6.14	El Bouni. Annaba	28
<i>V. sativa subsp. consobrina</i>	64	1.6.14	Azzaba. Skikda	111
<i>V. sativa subsp. consobrina</i>	86	13.6.14	Djbel el Ouehch Constantine	880
<i>V. sativa subsp. cordata</i>	13	26.5.14	El Kantra. Biskra	584
<i>V. sativa subsp. cordata</i>	42	28.5.14	Ain Abid. Constantine	847
<i>V. sativa subsp. cordata</i>	47	22.5.14	Frères Mentouri Constantine University	604
<i>V. sativa subsp. obovata</i>	10	27.5.14	Didouche Mourad. Constantine	468
<i>V. sativa subsp. obovata</i>	17	22.5.14	Chaab Ersas. Constantine	563
<i>V. sativa subsp. obovata</i>	22	2.6.14	Sigus. Oum El Bouaghi	822
<i>V. sativa subsp. obovata</i>	32	22.5.14	Chaab Ersas. Constantine	562
<i>V. narbonensis</i>	23	30.5.14	Hamma Bouziane. Constantine	425
<i>V. narbonensis</i>	55	27.5.14	Didouche Mourad. Constantine	443
<i>V. monantha subsp. calcarata</i>	18	3.6.14	Ain Taghrout. Bourdj Bou Areridj	934
<i>V. monantha subsp. calcarata</i>	78	20.5.14	Coudiat. Constantine	633
<i>V. monantha subsp. calcarata</i>	102		Frères Mentouri Constantine University	604
<i>V. leucantha</i>	100	10.6.14	INATAA. Constantine	586
<i>V. tenuifolia</i>	56	6.6.14	Ain Temouchent	276
<i>V. tenuifolia</i>	89	6.6.14	Sidi Khaled. Sidi Bel Abbes	543
<i>V. lutea subsp. vestita</i>	4	22.5.14	Frères Mentouri Constantine University	604
<i>V. lutea subsp. eu-lutea</i>	87	30.5.14	El Milia. Jijel	28

Extraction of the seed oils

The extraction of the seed oils was done according to Hara and Radin (1978) and to Emre *et al.* (2013), modified. The extraction of oils was done by Soxhlet (ISO-LAB) adopting the following approach: between 2 and 3 grams of finely ground seeds are deposited in cellulose extraction thimble (FILTER.LAB Barcelona, Spain). A piece of cotton is placed on it to prevent overflow of the sample and then the thimble is placed in a 250 ml Soxhlet flask. For each sample, the oil is extracted with 350 ml of n-hexane. The latter is brought to a boil then the temperature is lowered. To prevent the system from heating, its temperature is maintained at 8° C with a cryostat (LAUDA). The 500 ml flask containing the sample is recovered after 6 hours, then, it is passing to the rotary evaporator (IKA RV 10) at a temperature of 41° C and 80 rpm, connected to a vacuum pump (V-700 BUCHI). When remaining between 25 and 30 ml of hexane, the sample is recovered in a 50 ml balloon and then passed again in the rotavap. to remove all hexane.

Oil yield

The oil yield is then calculated by relating the weight of the extract to the weight of crushed seeds. The result was expressed as the lipid percentage in the dry seed powder.

Oil transesterification

The FAME were prepared by transesterification by mixing n-hexane with the oil then adding KOH (2 mol/L), according to Alves *et al.* (2008). After centrifugation, the sample is left in the dark for 1 hour and a half.

Fatty acids Methyl Esters analyses (FAME)

The FAME were analyzed by GC-FID, according to Renna *et al.* (2014), modified, to determine FA constituents. A 1 mL sample from the supernatant was put into GC vials and injection was started immediately. A gas chromatography system (GC-2010 plus SHIMADZU) equipped with a flame ionization detector and a TR-CN 100 column (100 m x 0.25 mm x 0.20 µm ID) was used. Injection block temperature was set at 250°C. The oven temperature was kept at 140°C for 5 min, then ramped up from 140 to 240°C at 4°C/min, and finally held at 240°C for 20 min. The injector (AOC-20i) is automated (0.80 µl per sample). Helium was used as carrier gas (flow rate 51.1 ml/min, split ratio 1/50). FAME peaks were identified by comparing their retention times and spectra with standard mixture analyzed under the same conditions. FA composition was expressed as % of total oil.

Cluster analysis based on fatty acids composition

Cluster analysis based on FA composition was performed with Euclidean Distances Matrix based on the Unweighted Pair-Group Method using Arithmetic Averages (UPGMA) by Statistica 6.0 software.

Results

Oil yields

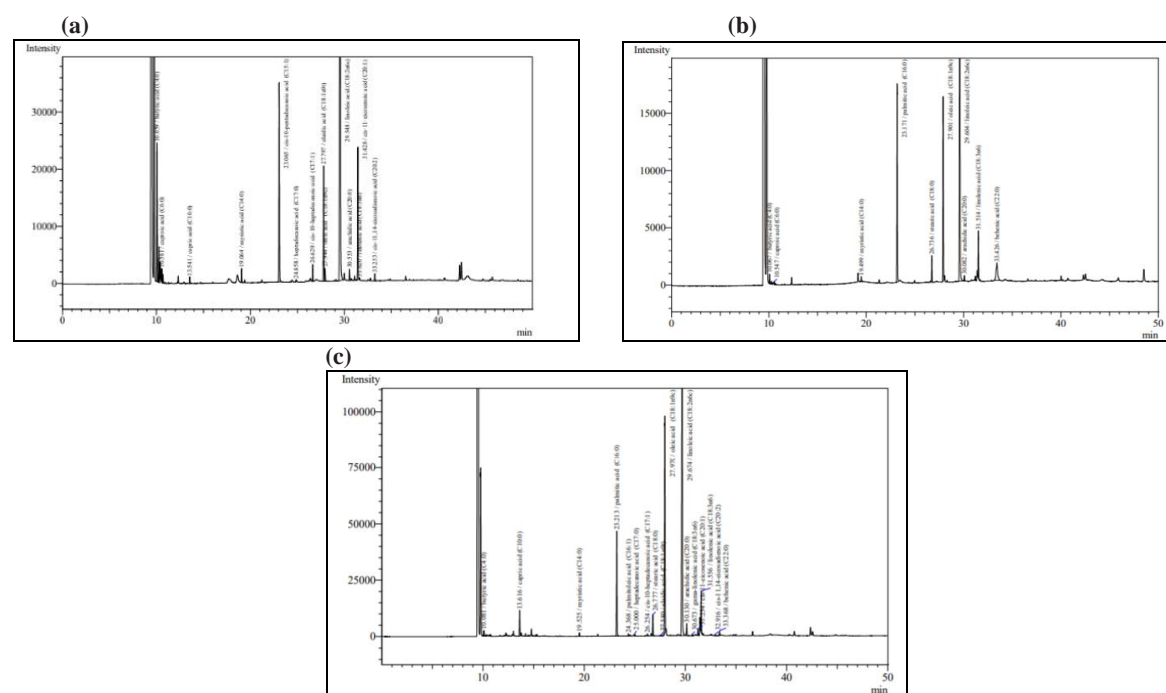
The oil yield of *Vicia* seeds varies from 0.67 to 2.40% (table 2). In the *sativa* complex, of the ten genotypes studied, an accession *obovata* has the highest oil yield (accession 22). The lowest yield is observed in an accession of *obovata* also. In the three accessions of *V. monantha* subsp. *calcarata*, a difference is observed in the oil yield of the three accessions. The only accession of *V. leucantha* is among the richest oil samples with a yield (2.16%). While, we recorded similar oil yield in both *V. tenuifolia* accessions.

Table 2. Seeds oils yields in accessions investigated

Taxon	Code	Oil yield (%)
<i>V. sativa</i> subsp. <i>consobrina</i>	14	1.35
<i>V. sativa</i> subsp. <i>consobrina</i>	64	1.50
<i>V. sativa</i> subsp. <i>consobrina</i>	86	2.23
<i>V. sativa</i> subsp. <i>cordata</i>	13	1.73
<i>V. sativa</i> subsp. <i>cordata</i>	42	1.8
<i>V. sativa</i> subsp. <i>cordata</i>	47	1.77
<i>V. sativa</i> subsp. <i>obovata</i>	10	1.53
<i>V. sativa</i> subsp. <i>obovata</i>	17	1.57
<i>V. sativa</i> subsp. <i>obovata</i>	22	2.40
<i>V. sativa</i> subsp. <i>obovata</i>	32	1.03
<i>V. narbonensis</i>	23	1.53
<i>V. narbonensis</i>	55	1.30
<i>V. monantha</i> subsp. <i>calcarata</i>	18	2.32
<i>V. monantha</i> subsp. <i>calcarata</i>	78	0.67
<i>V. monantha</i> subsp. <i>calcarata</i>	102	1.30
<i>V. leucantha</i>	100	2.16
<i>V. tenuifolia</i>	56	1.24
<i>V. tenuifolia</i>	89	1.2
<i>V. lutea</i> subsp. <i>vestita</i>	4	1.33
<i>V. lutea</i> subsp. <i>eu-lutea</i>	87	1.13

Fatty acids analysis

Chromatograms of some accessions are presented in figure 1. A total of 27 different FA were identified (table 3), varying from 9 to 20 FA.


Fig 1. Chromatograms of some accessions obtained by GC-FID of fatty acids

(a) : accession 100 (b) accession 18 (c) accession 55

Table 3. Fatty acids composition of *Vicia* taxa studied

FA (%)	14	64	86	13	42	47	10	17	22	32	23	55	18	78	102	100	56	89	4	87
Butyric acid	3.91	4.70	4.81	7.722	2.84	-	0.94	0.078	0.265	0.227	0.406	0.464	0.698	-	0.537	5.975	0.168	-	0.104	-
Caproic acid	-	-	-	2.312	0.359	0.096	-	-	-	-	-	-	0.294	-	0.132	0.567	0.098	0.161	-	0.129
Caprylic acid	-	-	0.311	-	-	-	-	-	-	-	-	-	-	0.255	-	-	-	-	-	-
Capric acid	0.352	1.802	3.593	2.932	-	-	-	-	-	-	-	2.526	-	-	-	0.331	-	-	-	-
Undecanoic acid	-	-	1.131	0.81	-	0.082	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Myristic acid	0.917	0.176	-	0.17	-	0.223	0.602	0.423	0.328	0.8	0.523	0.284	0.403	1.637	0.471	0.68	0.909	1.213	0.493	1.048
Pentadecanoic acid	0.249	-	-	0.238	-	0.239	0.194	0.168	0.156	0.231	0.123	-	-	0.268	0.263	-	0.359	0.317	0.246	0.27
Cis-10 pentadecanoic acid	-	-	14.126	-	-	-	-	-	-	-	-	-	-	-	-	9.51	-	-	-	-
Palmitic acid	11.737	12.457	-	10.73	12.902	12.739	13.913	11.028	13.445	11.913	10.677	10.41	15.075	14.525	13.308	-	13.065	14.369	11.563	11.932
Palmitoleic acid	-	0.233	-	0.06	0.077	-	-	0.075	0.063	-	0.135	0.212	-	-	-	-	0.078	-	0.075	-
Heptadecanoic acid	0.19	0.246	-	0.149	0.151	0.183	0.169	0.162	0.153	0.169	0.19	0.18	-	-	0.212	0.082	0.217	0.249	0.225	0.154
Cis-10 heptadecanoic acid	0.145	-	4.023	0.052	0.1	0.104	-	-	-	0.127	-	0.172	-	-	-	0.776	0.208	-	-	-
Stearic acid	3.188	4.455	-	2.675	2.469	3.607	2.798	3.309	2.835	2.429	2.702	2.124	1.994	1.637	1.561	-	1.939	2.296	2.644	2.271
Elaidic acid	-	0.265	19.507	-	-	-	-	-	-	-	-	0.155	-	-	-	0.596	-	-	-	-
Oleic acid	17.102	16.497	0.071	14.016	19.396	19.572	14.522	21.835	14.279	16.35	35.882	24.866	14.17	12.87	16.05	0.647	20.072	18.449	15.495	9.989
Linolelaidic acid	-	0.169	0.894	0.064	-	-	0.068	-	0.064	-	-	-	-	-	-	-	-	-	-	-
Linoleic acid	51.554	48.884	46.746	45.282	51.653	50.364	56.344	51.659	56.982	55.728	41.16	51.756	58.77	62.948	60.734	67.911	50.206	48.024	60.299	61.764
Linolenic acid	8.055	4.866	-	8.578	8.019	9.073	8.43	7.876	9.774	9.384	3.614	4.032	3.799	-	4.454	0.078	10.317	11.984	7.344	9.499
Arachidic acid	0.897	-	0.282	1.234	1.136	1.852	1.217	1.395	1.313	1.108	1.151	1.379	0.461	0.666	0.738	0.617	0.811	1.323	0.934	0.975
Gamma-linolenic acid	-	-	-	0.297	0.178	0.344	0.11	-	-	0.151	0.083	0.14	-	5.194	0.35	-	0.44	0.587	-	0.253
Heneicosanoic acid	-	-	0.816	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Homolinoleic acid	1.322	0.308	-	-	0.283	-	0.308	0.982	-	0.976	-	0.119	-	-	0.542	0.349	0.649	0.465	0.258	1.386
Cis-11, 14, 17-ecosatrienoic acid	-	-	-	-	-	1.049	-	0.133	-	-	-	-	-	-	-	-	-	0.203	-	-
Cis-13, 16-docosadienoic acid	-	-	-	0.379	-	-	-	0.455	-	-	0.368	-	-	-	-	-	-	0.359	-	-
Cis-11-eicosenoic acid	0.381	3.708	3.694	1.942	0.444	0.475	0.384	0.422	0.345	0.409	0.593	0.67	-	-	0.647	6.882	0.464	-	0.321	0.329
Behenic acid	-	0.284	-	0.358	-	-	-	-	-	-	-	0.51	4.336	-	-	-	-	-	-	-
Cis-4, 7, 10, 13, 16, 19-docosaheptaenoic acid	-	-	-	-	-	-	-	-	-	-	2.393	-	-	-	-	-	-	-	-	-
Number of fatty acids	14	16	13	20	14	15	14	15	13	14	15	17	10	9	14	14	16	14	13	13
Σ SFA	21.44	24.12	10.95	29.33	19.857	19.021	19.833	16.563	18.495	16.877	15.649	17.877	23.261	18.988	17.222	8.601	17.349	19.928	16.209	16.779
Σ MUFA	17.628	20.703	41.421	16.07	20.017	20.151	14.906	22.332	14.687	16.886	36.61	26.075	14.17	12.87	16.697	23.411	20.822	18.449	15.891	10.318
Σ PUFA	60.931	54.227	47.64	54.6	60.133	60.83	65.26	61.105	66.82	66.239	47.618	56.047	62.569	68.142	66.08	68.338	61.612	61.622	67.901	72.902
Σ UFA	78.559	74.930	89.061	70.67	80.150	80.981	80.166	83.437	81.507	83.125	84.228	82.122	76.739	81.012	82.777	91.749	82.434	80.071	83.792	83.22
USFA / SFA	3.66	3.11	8.13	2.41	4.04	4.26	4.04	5.04	4.41	4.93	5.38	4.59	3.30	4.27	4.81	10.67	4.75	4.02	5.17	4.96
ω6	52.876	49.361	47.64	46.022	52.114	50.708	56.83	53.096	57.046	55.879	41.611	52.015	58.77	68.142	61.084	68.26	51.295	48.97	60.557	63.403
ω3	8.055	4.866	0	8.578	8.019	10.122	8.43	8.009	9.774	9.384	6.007	4.032	3.799	-	4.454	0.078	10.317	12.187	7.344	9.499
ω6 / ω3	6.564	10.144		5.365	6.499	5.010	6.741	6.630	5.837	5.955	6.927	12.901	15.470		13.714		4.972	4.018	8.246	6.675

Saturated fatty acids

The SFA content varied from 8.601 (*V. leucantha*) and 29.33% (*V. sativa* subsp. *cordata*), with the predominance of palmitic acid which was observed at the highest level in an accession of *V. monantha* subsp. *calcarata* and at the lowest amount in an accession of *V. narbonensis*. The highest rate of butyric acid is recorded in a *V. sativa* subsp. *cordata* sample, the lowest rate is also found in the *sativa* complex but in another taxon. This FA was not identified in 4 accessions. It should be noted that the caproic acid was not detected in 11 accessions out of the 20 studied. Its highest and lowest rates are registered in samples of *V. sativa* subsp. *cordata*.

Caprylic acid has been identified in only two accessions, as for the capric acid, it was recorded in 6 samples with the highest rate in a population of *V. sativa* subsp. *consobrina*. Special attention can be attributed to undecanoic acid, which was detected only in a single accession (*V. sativa*, subsp. *consobrina*), with a rate of 1.131%. As for myristic acid, the highest rate of undecanoic acid is detected in a genotype of *V. monantha* subsp. *calcarata*, its lowest one in a genotype of *V. narbonensis*. Accordingly, saturated acids with low molecular weight (caproic, caprylic, capric) can be found in certain accessions; but not in others.

Note that heneicosanoic acid is only found in the sample 86 (*V. sativa* subsp. *consobrina*). Behenic acid was found in only 4 samples with the highest level in an accession of *V. monantha* subsp. *calcarata*.

Total UnSaturated fatty acids (TUSFA)

The TUSFA content is much higher than that of SFA. It varies from 70.67 to 91.749%, registered in the single accession of *V. leucantha* (table 3).

MonounSaturated fatty acids (MUSFA)

Six MUFA were detected out of 27 obtained. The lowest level is observed in an accession of *V. lutea* subsp. *eu-lutea*; the highest rate is detected in a sample of *V. sativa* subsp. *consobrina*.

The cis-10 pentadecanoic acid is recorded only in a *V. sativa* subsp. *consobrina* genotype as well as in the only accession of *V. leucantha*. Palmitoleic acid is found in 10 genotypes with its highest and lowest levels detected in the complex *sativa* but in two different taxa, which is also valid for cis-10 heptadecanoic acid.

Elaidic acid occurs only in 3 genotypes with its highest level in an accession of *V. sativa* subsp. *consobrina*.

Special mention should be made for oleic acid, which is the most common MUSFA in the genotypes studied. Its highest rate was obtained in an accession of *V. narbonensis*. Finally, cis-11 eicosenoic acid is registered in 17 accessions, its highest level is detected in the single accession of *V. leucantha*. The cis-10-pentadecanoic acid is observed in only two samples: *V. sativa* subsp. *consobrina* and the single accession of *V. leucantha*.

Total Polyunsaturated fatty acids (TPUFA)

The TPUFA varies from 47,618 to 72,902% (table 3). The highest amount is found in a genotype of *V. lutea*, when the lowest one is obtained in a sample of *V. narbonensis*. Linolelaidic acid was only observed in 5 samples. Linoleic acid is the only FA found in all genotypes and is the most common among PUFA. It varies from 41.16 to 67.911% (in the only accession of *V. leucantha*). This accession has the lowest level of linolenic acid. γ -linolenic acid was not identified in 9 samples, its highest level was found in an accession of *V. monantha* subsp. *calcarata*.

Cis 11, 14, 17 ecosatrienoic acid was only identified in 3 accessions. Cis 13, 16-docosadienoic acid was observed in only 4 samples. Cis 4-7-10-13-16-19 docosaheptaenoic acid was only identified in a *V. narbonensis* accession. The cis-13,16-docosadienoic acid is observed only in four accessions.

Cluster analysis based on fatty acids composition

At the distance of 239.96, the dendrogram (figure 2), can be divided into two major clusters.

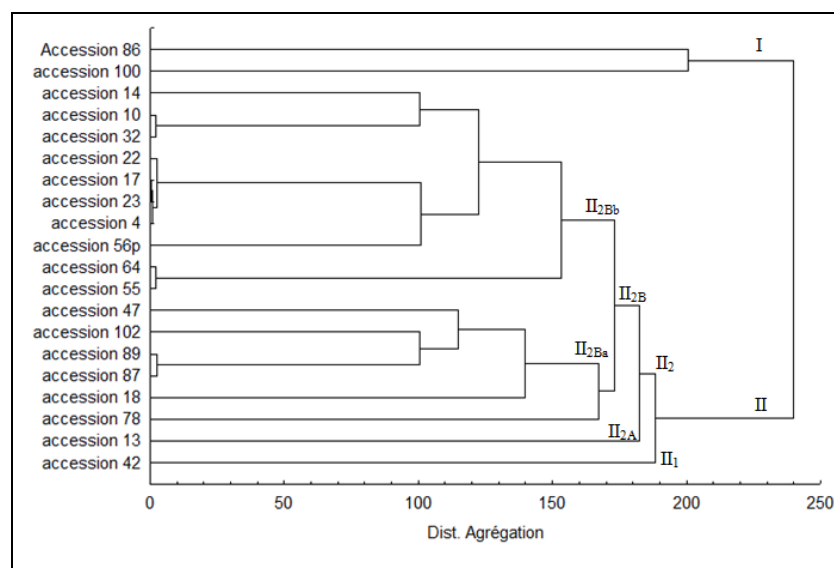


Figure 2. Cluster analysis based on fatty acids composition

In the subcluster II₂B_a, containing six samples, four of them belong to the subgenus *Vicilla*, section *Cracca*. The other two belong to the subgenus *Vicia*; a sample is part of the *Vicia* section and the second part of the *Hypechusa* section. In the subcluster II₂B_b, we find on one side two accessions belonging to the same subgenus and to the same section: *V. narbonensis* (Subgenus *Vicia*, section *Narbonensis*) and *V. sativa* subsp. *consobrina* (Subgenus *Vicia*, section *Vicia*). On the other side, three groups are defined: the first group contains three samples of the same species and therefore of the same genus, the same sub-genus and of the same section (subgenus *Vicia*, section *Vicia*), the second group contains an accession of the subgenus *Vicilla* and the section *Cracca*, the third group includes 4 accessions, the first of which belongs to the subgenus *Vicia*, section *Hypechusa* and the other three to the subgenus *Vicia* and to two different sections (*Vicia* and *Narbonensis*).

The largest distances corresponding to very distant samples in the clustering, is observed between two accessions belonging to the two subgenera (*Vicia* and *Vicilla*) and therefore to different sections and species. In parallel, the smallest distance is obtained between a *sativa* accession (subgenus *Vicia*, section *Vicia*) and an accession of *Narbonensis* (subgenus *Vicia*, section *Narbonensis*). At the same time, smaller distances are obtained between genotypes belonging to different sections (*Narbonensis*, *Vicia* and *Hypechusa*).

Discussion

The lowest and highest values of oil yields are observed in accessions belonging to different species. In addition, accessions belonging to the same taxon have different oil yields, as is the case with the three samples of *V. monantha* subsp. *calcarata*.

No literature exists reporting the FA profile of the different studied taxa. In addition, some FA identified in our accessions have not been found in previous publications. However, our results are in accordance with many works where USFA were higher than SFA, and PUSFA upper than MUSFA levels, as reported in the seeds of various legumes in different ecogeographical areas (Bagci *et al.* 2004 ; Pastor Cavada *et al.* 2009b; Renna *et al.* 2014). Moreover, our findings are in agreement with Bagci *et al.* (2004), who recorded that the main FA in legume seed are palmitic, oleic, linoleic and linolenic acids.

From the other hand, the ratio SFA/USFA is superior than those reported for Chinese (Akpınar *et al.*, 2001), and Turkish vetch (Mao *et al.*, 2015). Besides, higher ratios of SFA/USFA are observed in our samples compared to previous studies (Renna *et al.*, 2014). According to Mao *et al.* (2015), the seeds are low in lipid with a ratio of USFA/SFA up to 3.30–3.66, which means that the seeds have accumulated more USFA primarily as linoleic, oleic and α -linolenic acids.

The oleic/linoleic FA ratio was usually reported to be less than one in the seeds of many *Vicia* taxa or Leguminosae (Pastor Cavada *et al.* 2009b).

The lowest level of palmitic acid was observed in a *V. narbonensis* accession with the highest seed diameter (5.50 ± 0.42). The lowest level of linolenic acid was observed in the single accession of *V. leucantha* which has a mean diameter compared to other taxa (Bechkri *et al.* 2017a). Our findings concord to Holman (1981), who observed that the proportion of palmitic and linolenic acids diminishes, as the diameter of peas increases during growth period

Total Saturated Fatty Acids

SFA values obtained in our study are different to those reported by Bağcı *et al.* (2004). Our results join previous data where palmitic and stearic acids are major SFA (Akpınar *et al.* 2001, Pastor-Cavada *et al.* 2009b). But, their concentrations are lower than those of Kaplan *et al.* (2014). In our study, the lower value was observed in the only sample of *V. leucantha*. While for Renna *et al.* (2014), the seeds of *V. narbonensis* showed a significantly lower TSFA concentration.

Odd-chain SFA (pentadecanoic, heptadecanoic, non-decanoic acids) detected in our samples, were not reported previously. However, Lauric acid which is not detected in our accessions was reported by Akpınar *et al.* (2001).

Low molecular acids from the SFA were absent or present at trace levels in the studied vetch oils, while margaric and margaroleic acids were not detected in our samples.

The arachidic acid was higher than 1%, which is different from finding of Bağcı *et al.* (2004), where it was lower than 1%. These researchers also reported that the highest and lowest absolute concentrations of this FA were observed in *V. sativa* subsp. *amphicarpha* and *V. narbonensis* (which is not our case).

The content of myristic and pentadecanoic acids in common vetch seed in our study is lower than those of Chinese (Mao *et al.*, 2015), and Turkish samples (Akpınar *et al.* 2001).

According to Bağcı *et al.* (2004), lignoceric acid was not detected in the seeds of *V. narbonensis* and *V. villosa* when vetch seeds showed significant differences in the concentration of lignoceric acid which is not observed in our genotypes.

Palmitic acid is a steady lipid of several Leguminosae and is the most important SFA in vegetable oils and especially in *Vicia* seeds (Renna *et al.* 2014). These findings agree with our results since the most important SFA found in our samples is the palmitic acid. Its amount was relatively low comparing with the values reported by Renna *et al.* (2014) and by Andrzejewska *et al.* (2016). The latter reported that the level of this FA in lupine is much higher than that of vetch.

Our results are in agreement to many previous works which reported that the stearic acid is the second most abundant FA in vetch seeds (Akpınar *et al.* 2001, Bağcı *et al.* 2004, Pastor Cavada *et al.* 2009b, Emre *et al.* 2011, Renna *et al.* 2014). Their values are similar to those reported for the other Leguminosae (Bağcı *et al.* 2004).

In our study, the richest sample in stearic acid is an accession of *V. sativa* subsp. *consobrina*, which concurs to Emre *et al.* (2011), who noticed that the richest species in stearic acid is a subspecies of *V. sativa*. Besides, stearic acid was found in a low level in our samples, while it was more important in Turkish seeds (Akpınar *et al.* 2001).

On the other side, Bagci *et al.* (2004), reported that the highest and lowest concentrations of behenic acid were observed in *V. sativa* subsp. *amphicarpha* and *V. narbonensis*. While, in our results, it was only revealed in 4 samples and with rates lower than 1% (except for a *V. monantha* subsp. *calcarata* accession). This result highlights a nutritional value of the samples since several studies revealed that oils with high levels of long chain SFA such as behenic acid may be difficult for the digesting enzymes in animals (Akpinar *et al.* 2001). No work in the literature has reported the presence of caproic, caprylic and capric acids, whereas these FA have been detected in our taxa.

Total UnSaturated Fatty Acids (TUSFA)

Our results show that TUSFA contents of the seeds oils are higher than the TSFA, which are higher than those found by Kaplan *et al.* (2014). In our study, we noticed an interesting rate of 91.74%, which was not reported in the literature.

Indeed, according to Bagci *et al.* (2004), TUSFA content of *Vicia* species is lower than 70%. While Bakoglu *et al.* (2017), reported that TUSFA of vetch taxa can reach 80.67%. They also determined that TUSFA was included between 75.9 and 83.46%, in earlier works, on different Leguminosae (Bakoglu *et al.*, 2010).

Furthermore, TUSFA values were comparable to those previously reported for other taxa of the genus (Bagci *et al.* 2004; Pastor-Cavada *et al.* 2009b; Emre *et al.* 2011; Renna *et al.* 2014), even though some of these works did not consider the same taxa we studied. Our results are also consistent with works on related genera like *Lathyrus*, *Lens*, *Pisum* and *Lupine* (Bagci *et al.* 2004). Nevertheless, Akpinar *et al.* (2001) noticed that TUSFA content of vetch seeds, grown in Turkey, is under 60 %.

According to several studies, oils rich in oleic and linoleic acids are the most adaptable and also excellent edible oils. Oleic, linoleic and linolenic acids are major USFA in the present study. All the investigated taxa were richer in oleic and linoleic acids than in linolenic acid. Oleic and linoleic acids were determined to be the major USFA in some Leguminosae oil which is used as a food in some countries (Higuchi *et al.* 1982). Oleic acid is well represented being the third most abundant USFA, after linoleic and oleic acids, in all the samples here analyzed. Its highest rate was observed in an accession of *V. tenuifolia*, a result that has never been reported in the literature.

MonoUnSaturated Fatty Acids (MUSFA)

Oleic acid has a role in the prevention of cardiovascular risks. This FA was found to be the most abundant one in the vetch by Pastor Cavada *et al.* (2009b). Our findings join those of Emre *et al.* (2011), who revealed that the major MUSFA in *Astragalus* seeds of all taxa is oleic acid. The same result was found on vetch by Akpinar *et al.* (2001). Compared to all other detected FA, oleic acid showed the greatest differences among the studied seeds which also concord to the results of Renna *et al.* (2014), who noticed that the seeds of *V. sativa* contained significantly lower concentrations of oleic acid than *V. narbonensis*. In our results, the highest level of oleic acid was also noticed in an accession of *V. narbonensis*. In fact, according to previous studies conducted in Mediterranean regions (Bagci *et al.* 2004, Pastor Cavada *et al.* 2009b, Emre *et al.* 2011, Renna *et al.* 2014), the highest oleic acid content was recorded in *V. narbonensis*.

Palmitoleic acid did not show significant difference among the considered seeds, its lowest level is detected in a sample of *V. sativa* subsp. *cordata*. This is in accordance with Emre *et al.* (2011), who noticed that the lowest percentage of this FA is revealed in a subspecies of *V. sativa*.

The seeds showed intermediate amounts of eicosenoic acid. The highest value was found in the single *V. leucantha* sample, which was not reported in any previous publication except in a small amount, as related by Mao *et al.* (2015).

Erucic acid exerts negative effects on animal metabolism (Kuhnt *et al.* 2012). The presence of the erucic acid in vetch seeds was reported in some legumes, but not in vetch seed (Akpinar *et al.*, 2001; Bagci *et al.*, 2004). However, erucic acid was not detected either in the Tunisian vetch (Renna *et al.*

(2014), or in the present work. No previously published data, in vetch seeds, reported the presence of the nervonic acid (an erucic acid derivative), that we detected in our samples.

From the other hand, a diet high in omega-9 could help to reduce the risk of developing cardiovascular disorders. In addition, since the ratio of omega-6/omega-3 is too high in modern diets, this gives another advantage to omega-9s. In our results, the only representative FA in omega 9 is the cis-11-eicosenoic acid.

PolyUnsaturated Fatty Acids (PUSFA)

PUSFA have a role in the initial oxidative processes in which FA are transformed to carbohydrates. Conjugated linoleic acid (CLA) or other PUSFA have anti-carcinogenic, anti-thrombogenic and anti-atherogenic properties. It is possible to increase CLA by increasing linoleic and linolenic acids in the feed as CLA are formed through isomerization of linoleic acid (Grinari *et al.* 2000). According to Hawke (1973), most of FA, mainly consists of palmitic, stearic, oleic, linoleic and linolenic acids, located in the chloroplasts. Their concentration varies among different environments, primarily due to variation in PUSFA, of which linoleic and linolenic acids play an important role in maintaining membrane structure integrity (Lee and Cho 2012).

According to Meydani *et al.* (1991), linoleic acid is needed for a normal immune response. In the current study, linoleic acid amounts can reach 67.91%. Pastor-Cavada *et al.* (2009b), and Bağcı *et al.* (2004), reported that levels of linoleic acid reach 66.3% and 50% of TFA, respectively. Emre *et al.* (2011) demonstrated that linoleic acid content in vetch is greater than 45%, while Akpinar *et al.* (2001), detected lowest linoleic acid content in *Vicia* taxa. In the same time, our results are consistent with those of several authors according to which, linoleic acid was usually found to be the most abundant FA in vetches and other legumes (Bağcı *et al.* 2004; Pastor-Cavada *et al.* 2009b).

According to Renna *et al.* (2014), *V. sativa* subsp. *amphicarpa* and *V. villosa* present higher concentrations of linoleic acid compared to *V. narbonensis*, while intermediate values detected for the other seeds. Our results show that *V. monantha* subsp. *calcarata*, *V. lutea*, but especially *V. leucantha* present the most important values of linoleic acid.

Our results show that linoleic acid is the predominant PUSFA, followed by linolenic acid, in the investigated taxa. The same findings were reported on Astragals, lupine and vetch (Emre *et al.* 2011; Renna *et al.* 2014).

Concerning linolenic acid, Akpinar *et al.* (2001) determined values of 1.9-9.2%. Bağcı *et al.* (2004) reported that linolenic acid content in the legume seeds generally found to be lower than 10%. Linoleic and linolenic acids, the two major PUSFA detected in our taxa, are essential FA which cannot be synthesized *de novo* and are needed for optimum development and health. Linoleic acid was reported as the major FA in some *Vicia* taxa but more usually as the third most abundant USFA (after linoleic and oleic acids) in other vetch species; these results also occurred in our study.

In our results, linolenic acid was detected at low levels, comparing to linoleic and oleic acids. Oil should have a minimal amount of linolenic acid since the latter is considered as the first FA responsible for undesirable flavors in stored oils (Wolff and Kwolek 1971).

Several works reported the presence of α -linolenic acid in oil obtained from Leguminosae (Renna *et al.* 2014). In vetch species, the content of this FA can reach 21.98%, in Turkey and 9.77%, in Tunisia. Many vetches were recognized to contain less than 15% α -linolenic acid in their seeds (Akpinar *et al.* 2001, Bağcı *et al.* 2004, Pastor-Cavada *et al.* 2009b). Exceptions regarded few species or varieties (Pastor-Cavada *et al.* (2009b; Bağcı *et al.*, 2004). In our results, the presence of α -linolenic acid was not detected.

No previous work reported the presence of γ -linolenic acid, which is the most important FA observed in a sample of *V. monantha* subsp. *calcarata*, and detected in 12 accessions. This FA acid possesses a therapeutic value. The presence of linolelaidic acid did not carry over into the literature. However, it

has been detected in some of our samples. Other FA have been revealed as traces, as reported by Renna *et al.* (2014).

The highest value of n6/n3 was observed in an accession of *V. monantha* subsp. *calcarata* and the lowest one in an accession of *V. tenuifolia*, which also has the highest level of linolenic acid. The latter was generally found to be lower than 10% in the legume seed oils. This observation applies to our results since the 10% rate was only exceeded for one of the 20 studied.

Previous study reported the presence of arachidonic acid in vetch oil seed with relatively low and comparable concentrations (Renna *et al.* 2014); which is not the case in our accessions.

According to Bagci *et al.* (2004), eicosadienoic acid was detected only in one out of six analyzed species. While, Akpınar *et al.* (2001) detected a large variation in the levels of this FA with a higher value of eicosadienoic acid if than those noticed in our study and by Renna *et al.* (2014).

Special attention must be given to the cis-4, 7, 10, 13, 16, 19 docosahexaenoic acid which has not been reported in the literature but which was found in an accession of *V. narbonensis*. Other FA (cis-11, 14, 17-ecosatrienoic and cis-13, 16-docosadienoic acids) have not been reported in previous studies but have been observed as traces in our genotypes

Our results demonstrate the quality of the genotypes, which contain FA of great nutritional interest. Indeed, Omega 3-FA which rate can reach 12.187% in our taxa are PUSFA which have been associated with many health benefits. Among the omega-6s, only linoleic acid is described as "essential". The body uses omega-6 to develop highly USFA and eicosanoids series 1 and 2.

Omega 6/Omega3

The n6/n3 FA ratio is used to assess the nutritional value of lipids. An optimal n6/n3 FA ratio should vary between 1:1 and 4:1, but Western diets may reach ranges of 10:1 to 20:1 (Simopoulos 2011), which corresponds to the values found in some of our accessions.

Environment influence

Our study shows qualitative and quantitative differences between taxa and even between accessions of the same taxon. Much dissimilarity between our results and those of other works have been observed. Obviously, several factors, such as genetics, geographical location, climatic settings and growing conditions may affect the FA content in the seed oils. Akpınar *et al.* (2001) reported that variations in the ecogeographical zones may also have exerted a key role as the environment can significantly affect the synthesis of FA. Murcia and Rincon (1992) reported that FA composition is influenced by agro-climatic conditions as the enzymes in FA biosynthesis depend on these factors. Environmental-based factors could contribute significantly to the observed differences as our accessions were collected from different regions.

According to Mao (2012), highest lipid and USFA contents were found in taxa characterized by lower accumulated temperature ($\geq 10^{\circ}\text{C}$) in the region. In our results, the highest lipid and USFA contents were observed in genotypes collected from stations characterized by cool winter. Plants increase the contents of lipids and USFA as a response to the drop in temperatures. During the process of cold accumulation, FA desaturases convert stearic acid to linolenic acid through oleic acid and linoleic acid intermediates (Falcone *et al.* 2004).

The higher content of SFA was observed in a sample collected in a station with warm winter. According to Lee and Cho (2012), the high temperature environment induces a rise of SFA which could be explained by lipid peroxidation accompanied by a simultaneous decrease of USFA. Similarly, Larkindale and Huang (2004) reported that the increase in SFA could enhance membrane rigidity, thus maintain membrane integrity under the high temperature environment.

Phylogenetic relationships

Bagci *et al.* (2004) reported that FA profiles can be used as a good chemotaxonomic marker for Fabaceae in general and for the genus *Vicia* in particular. The FA profiles obtained highlighted both inter and intraspecific variability. Apart from a few isolated cases, genotypes of the subgenus *Vicilla* and the section *Cracca* tend to cluster together. Similarly, accessions of the subgenus *Vicia* also tend to cluster together.

These results correspond to those of our previous works (Bechkri and Khelifi 2016, Bechkri and Khelifi 2017 a, b, Bechkri *et al.* 2018, Bechkri *et al.* 2019), where the separations were sufficient on the sectional level and where the species of *sativa* exhibited a set of taxa with an overlap. According to Abozei *et al.* (2017), all species from section *Cracca* clustered together, using FA profiles showing a close relationship among the section members.

It is very important to emphasize that the accessions of the *Narbonensis* section (represented by *V. narbonensis*) bind to the accessions of the *Vicia* section (represented by *V. sativa*). This result is also reported by Schaefer *et al.* (2012).

We highlight the particularity of *V. leucantha*, which is far from the other accessions because of its particular content in FA. It also has the highest level of linoleic acid, as well as the highest USFA/SFA ratio. These results were not obtained by previous works. This genotype is among the samples that show a high oil yield compared to the others. According to Liu (2011), the differences in changing intensity of FA composition among grain species correspond to those in oil distribution in the seed, while a varietal difference in distribution patterns and the FA composition of lipids within the species were insignificant.

It is important to mention that the accessions of the first cluster belong to the same bioclimatic stage. But in the other clusters, the genotypes that are found together belong to different bioclimatic stages (Bechkri and khelifi 2016, 2017a, 2017b). At the same time, the smallest clustering distance was obtained between samples collected from the same city, characterized by a cool winter. Thus there is no clear relationship between the grouping of samples according to their content in FA and their geographical origin.

Conclusion

In this study, vetch seed oils were rich in palmitic, stearic, oleic, linoleic and linolenic acids. Despite the small proportion of lipids present in legumes seeds, their profiles indicate the desirable nature of FA present. It is very likely that this content is influenced by the land type, altitude and an average temperature in the growing location. Obtained results might provide contributions on the chemotaxonomic and phylogenetic relationships. However, due to the high level of crossbreeding in vetch taxa, the FA profile cannot be used as a tool for taxonomic determination at specific level, but, it reflects the subgeneric and sectional affinities. The evaluation of FA in a wider range of *Vicia* taxa is suggested to characterize the chemotaxonomic and phylogenetic relationships both on intra and interspecific levels, especially in *sativa* complex. The possible toxic effect should also be kept in mind and other compounds related to nutritional quality should be investigated.

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